

**Turkish Journal of Agriculture - Food Science and Technology** 

Available online, ISSN: 2148-127X | www.agrifoodscience.com | Turkish Science and Technology Publishing (TURSTEP)

# Molecular Survey of the *Toxoplasma gondii* and *Neospora caninum* in brain tissue of aborted fetuses of Morkaraman sheep in Muş, Türkiye

Davut Koca<sup>1,a,\*</sup>, Burçak Aslan Çelik<sup>2,b</sup>, Özgür Yaşar Çelik<sup>3,c</sup>, Adnan Ayan<sup>4,d</sup>, Özlem Orunç Kılınç<sup>5,e</sup>, Ali Osman Turgut<sup>6,f</sup>, Özge Oktay Ayan<sup>7,g</sup>

<sup>1</sup>Van Yuzuncu Yil University, Faculty of Veterinary Medicine, Department of Obstetrics and Gynecology, Van, Türkiye
 <sup>2</sup>Siirt University, Faculty of Veterinary Medicine, Department of Parasitology, Siirt, Türkiye
 <sup>3</sup>Siirt University, Faculty of Veterinary Medicine, Department of Internal Medicine, Siirt, Türkiye
 <sup>4</sup>Van Yuzuncu Yil University, Faculty of Veterinary Medicine, Department of Genetics, Van, Türkiye
 <sup>5</sup>Van Yuzuncu Yil University, Özalp Vocational School, Department of Medical Laboratory Technician, Van, Türkiye
 <sup>6</sup>Siirt University, Faculty of Veterinary Medicine, Department of Animal Science, Siirt, Türkiye
 <sup>7</sup>Van Yuzuncu Yil University, Faculty of Medicine, Department of Parasitology, Van, Türkiye
 <sup>\*</sup>Corresponding author

ARTICLE INFO	A B S T R A C T
Research Article	<i>Toxoplasma gondii</i> and <i>Neospora caninum</i> are obligate intracellular protozoan parasites that can affect different warm-blooded species worldwide. In this study, it was aimed to detect <i>T. gondii</i> and <i>N. caninum</i> using PCR method in brain tissues of aborted sheep fetures. Brain specimens were
Received : 17.10.2023 Accepted : 21.11.2023	collected from 50 Morkaraman sheep fetuses that had undergone abortion at various stages of pregnancy, within the lambing seasons of 2023 in Muş. Approximately 1 cm <sup>3</sup> of brain tissue from
<i>Keywords:</i> Abortion <i>Toxoplasma gondii</i> <i>Neospora caninum</i> PCR Sheep	the right cerebral hemisphere was excised and subsequently frozen at -20°C for DNA extraction. DNA extraction and PCR amplification were then performed. As a result of this study, 11 (22%) of 50 brain tissues were positive. All brain samples examined in this study were negative for <i>Neospora</i> <i>caninum</i> . Based on the results of this study, it is possible to say that <i>T. gondii</i> is an important abortion agent in sheep in this region. Although <i>N. caninum</i> was not detected in this study, larger scale studies are recommended. Moreover, this study provides important information to breeders and veterinarians in the evaluation and management of abortion in the field.
<sup>a</sup> a davutkoca@yyu.edu.tr b http://www.edu.tr	ps://orcid.org/0000-0002-7962-6959 burcakaslan@siirt.edu.tr  ps://orcid.org/0000-0002-7962-6959 burcakaslan@siirt.edu.tr  ps://orcid.org/0000-0002-6564-3416 bookstanter  book



 $\odot \odot \odot$ 

This work is licensed under Creative Commons Attribution 4.0 International License

## Introduction

*Toxoplasma gondii* and *Neospora caninum* are obligate intracellular protozoan parasites that are distributed worldwide, have structural, genetic and immunological similarities, belong to the phylum Apicomplexa and can affect different warm-blooded species (Dubey, 2003; Basso et al., 2022).

*Toxoplasma gondii* causes toxoplasmosis, one of the most common parasitic diseases in humans and animals worldwide (Tenter et al., 2000; Wang et al., 2011). The definitive hosts are cats and felidae family, and the intermediate hosts are mammals, including humans, and birds (Dubey, 1994; Mor and Arslan, 2007). Infected cats can shed millions of oocysts. This causes widespread environmental contamination and is an important source of

infection for herbivores (Innes et al., 2009; Can, 2010). The infective forms of the parasite are tachyzoites, bradyzoites and oocysts of the felidae family, especially in the brain and muscle tissue. (Muz et al., 2013). The disease causes significant economic losses especially in sheep due to pneumonia, enteritis, neurological disorders, encephalitis, premature birth, stillbirth, neonatal losses and abortions (Van der Puije et al., 2000; Bártová et al., 2009; Dubey, 2009; Anğ et al., 2011).

*Neospora caninum* is an obligate intracellular parasite first isolated in Norway in 1984 in puppies with congenital encephalomyelitis (Dubey et al., 2007; Uzêda et al., 2007). The definitive host of *N.caninum* is domestic and wild canids and the intermediate hosts are herbivores.

Herbivores become infected by ingesting infected oocysts scattered in the feces of the definitive hosts (Sharma et al., 2015; Gharekhani et al., 2016). This disease causes abortions and newborn deaths in cattle, sheep and goats (Dubey, 2003; Figliuolo et al., 2004; Uzêda et al., 2007). In many hosts, transmission is transplacental. Significant economic losses can occur due to abortions and neonatal mortality (Sharma et al., 2015). In experimental studies, congenital infection has been reported in sheep and goats (Gharekhani et al., 2016).

In this study, it was aimed to detect *T. gondii* and *N. caninum* using PCR method in brain tissues of 50 aborted sheep foetuses in the Muş, Türkiye.

# **Material and Methods**

# The Study Area and Samples Collection

Brain specimens were collected from 50 sheep fetuses that had undergone abortion at various stages of pregnancy, within the lambing seasons of 2023 in Muş, Türkiye. The samples were exclusively sourced from Morkaraman breed sheep. Out of 50 aborted ovine fetuses, a total of 50 brain tissue samples were procured. To extract brain samples, each fetus was handled individually, with the calvarium opened and meninges dissected using a fresh disposable scalpel and forceps. Approximately 1 cm<sup>3</sup> of brain tissue from the right cerebral hemisphere was excised and subsequently frozen at -20°C for DNA extraction.

# **DNA** Extraction

DNA extraction was performed in all aborted fetus brain using the PureLink<sup>TM</sup> Genomic DNA Mini Kit (Invitrogen<sup>TM</sup>, USA, K182002), according to the manufacturer's protocol. The obtained DNAs were stored at  $-20^{\circ}$ C.

#### **PCR** Amplification

The amplification of the 529-bp repetitive element region of *Toxoplasma gondii* was conducted using the TgTox4F (5'-CGCTGCAGGGAGGAAGACGAAAGTTG-3') and TgTox4R (5'-CGCTGCAGACACAGTGCATCTGGATT-3') primers (Sah et al., 2019). Protocol for reactions was performed according to Orunç Kılınç et al. (2023).

For amplification of the Nc5 gene region of *N. caninum*, nested PCR was performed using external (5'-CTGCTGACGTGTCGTTGTTG-3') forward and (5'CATCTACCAGGCCGCTCTTC-3') reverse primers inner (5'-GCGTCAGGGTGAGGACAGTG-3') forward and (5'-CTCTCCGTTCGCCAGCAGTG-3') reverse primers (Fish et al., 2007). Protocol for both reactions was performed according to Orunç Kılınç et al. (2023).

The reaction was performed in an automatic thermal cycler (Eppendorf Mastercycler® pro) device. Subsequently, 1.5% agarose gel was prepared and stained with RedSafe<sup>™</sup> Nucleic Acid Staining Solution. The PCR products were run on an agarose gel afterward, and images were obtained on the gel imaging device (Syngene bioimaging system).

### Ethical Approval

This study was approved by Van Yuzuncu Yil University Animal Experiments Local Ethics Committee (Approval no:2023/03-10).

#### Results

In this study, a total of 50 brain tissue samples were chosen from aborted fetuses for the isolation of the *Toxoplasma gondii* parasite through conventional PCR. Positivity for the presence of *T. gondii* was confirmed in 11 out of the 50 samples, accounting for 22% of the total. (Figure 1). All brain samples examined in this study were negative for *Neospora caninum*.



Figure 1. Amplification of *T.gondii* using conventional-PCR. Lanes M: Marker, N: Negative control, P: positive control; 36, 28, 12 and 4 represent *T.gondii*. (529 bp).

#### Discussion

Toxoplasmosis plays an important role in sheep abortions and causes heavy losses to the sheep industry worldwide (Dubey, 2009). Especially infectious abortions have important effects on public health not only with their economic dimension but also with their zoonotic importance (Har and Başbuğan, 2019). Although *N. caninum* has been reported to cause congenital infections, abortions and deaths in newborn lambs in sheep, it is not considered among the main causes of abortion in sheep (Innes et al., 2001; Koyama et al., 2001; Hässig et al., 2003).

Toxoplasma gondii infections have been recorded in sheep populations worldwide with highly variable seroprevalences (Basso et al., 2022). A prevalence of 51.76% was reported in Egypt (Ibrahim et al., 2017), 38.22% in Brazil (Ueno et al., 2009), 3.76% in India (Sharma et al., 2008) and 1.6% in Iran (Raeghi et al., 2011). In studies conducted in Turkey; 54.65% prevalence was reported in Afyonkarahisar (Çiçek et al., 2004), 66.66% in Yalova (Oncel et al., 2005), 10% in Nevşehir (Çakmak and Karatepe, 2017), 48.4% in Mersin (Öztürk et al., 2002), 95.7% in Kars (Mor and Arslan, 2007) and 13% in Konya (Aköz et al., 2009). It was reported that T. gondii DNA was detected in 3 of 20 aborted fetal brain tissues (Hässig et al., 2003), 4 of 74 sheep fetal brain tissues (Moreno et al., 2012), 48 of 75 sheep fetal brains (Shahbazi et al., 2019), 9 of 111 sheep fetal brains (Partoandazanpoor et al., 2020), and 9 of 53 sheep fetal brains (Hurtado et al., 2001). In this study, the brains of 50 aborted sheep fetuses were examined by conventional PCR method and Toxoplasma

*gondii* DNA was detected in 11 (22%) tissues. This result was lower than the findings of some researchers (Öztürk et al., 2002; Çiçek et al., 2004; Oncel et al., 2005; Ueno et al., 2009; Ibrahim et al., 2017) and higher than the results of some studies (Aköz et al., 2009; Çakmak and Karatepe, 2017).

In studies to determine the prevalence of *N.caninum* in sheep in the world; 10.1% in Spain (Panadero et al., 2010), 16.8% in Greece (Anastasia et al., 2013), 8.81% in Brazil (Ueno et al., 2009), 27.7% in Pakistan (Nasir et al., 2012) and 1.53% in Iran (Ezatpour et al., 2015) prevalence was reported. In Türkiye, studies investigating the prevalence of *N. caninum* in sheep are quite limited. Positive rates of 12.4% in Adana (Ekşi et al., 2018), 0% in Van (Har and Başbuğan, 2019), 0% in Elazığ (Özkaraca et al., 2016) and 2.1% in Kars (Gökçe et al., 2015).

In studies conducted on aborted fetal brain tissue, it was reported that *N. caninum* DNA was detected in 4 of 20 aborted fetal brain tissues (Hässig et al., 2003), 5 of 74 sheep fetal brain tissues (Moreno et al., 2012), 3 of 18 fetuses (Howe et al., 2008), 18.9% of 74 fetuses (Hughes et al., 2006). All tissue samples examined in this study were negative for *Neospora caninum*. This result is similar to the findings of researchers (Özkaraca et al., 2016; Har and Başbuğan, 2019). The reasons for the differences between the studies include geographical location, climate, sheep breed, end-host prevalence and the tests used.

## Conclusion

Based on the results of this study, it is possible to say that T. gondii is an important abortion agent in sheep in this region. It was also concluded that PCR method is an important tool in the diagnosis of protozoal abortion agents. Although N. caninum was not detected in this study, larger scale studies are recommended. Furthermore, the results of this study are poised to have a significant impact on increasing awareness among veterinarians, researchers, and farmers regarding the epidemiology and prevalence of T. gondii and N. caninum in the Mus region. Nonetheless, additional investigations would he advantageous in elucidating the diverse genotypes of T. gondii and their potential correlation with abortion and other reproductive challenges in the sheep population. This will make a substantial contribution towards a more thorough comprehension of the influence of these factors on pertinent animals.

### **Conflicts of interest**

Authors state no conflict of interest.

#### References

- Aköz M, Aydin İ, Kamburgil K, and Handemir E. 2009. Determination of *Toxoplasma gondii* Seroprevalence by Indirect Fluoreseent Antibody (IFA) Test in Abortion Experienced and Abortion Inexperienced sheep in Karapınar District of Konya. Vet Bil Derg 25: 37-43.
- Anastasia D, Elias P, Nikolaos P, Charilaos K, and Nektarios G. 2013. Toxoplasma gondii and Neospora caninum seroprevalence in dairy sheep and goats mixed stock farming. Vet parasitol 198: 387-390.

- Anğ Ö, Tümbay E, and Küçüker M. (2011). Zoononoses: Infectious Diseases Transmissible from Animal to Human. İstanbul: Nobel Tıp Kitabevi.
- Bártová E, Sedlák K, and Literák I. 2009. *Toxoplasma gondii* and *Neospora caninum* antibodies in sheep in the Czech Republic. Veterinary Parasitology 161: 131-132.
- Basso W, Holenweger F, Schares G, Müller N, Campero LM, Ardüser F, Moore-Jones G, Frey CF, and Zanolari P. 2022. *Toxoplasma gondii* and *Neospora caninum* infections in sheep and goats in Switzerland: Seroprevalence and occurrence in aborted foetuses. Food and waterborne parasitology 28: e00176.
- Can M. 2010. The Assessment of Toxoplasmosis in Small Ruminants based on the Animal Health Economics Perspective. Atatürk Üniversitesi Vet Bil Derg 5: 167-174.
- Çakmak DÖ, and Karatepe B. 2017. Seroprevalence of *Toxoplasma gondii* in sheep from Nevşehir province in Turkey. Türkiye Parazitol Derg 41: 148-151.
- Çiçek H, Babür C, and Karaer Z. 2004. Seroprevalence of *Toxoplasma gondii* in sheep using Sabin-Feldman (SF) dye test in Afyon province. Ankara Univ Vet Fak Derg 51: 229-231.
- Dubey J. 1994. Toxoplasmosis. JAVMA 205: 1593-1597.
- Dubey J. 2003. Review of *Neospora caninum* and neosporosis in animals. Korean J. Parasitol 41: 1-16.
- Dubey J. 2009. Toxoplasmosis in sheep-the last 20 years. Vet parasitol 163: 1-14.
- Dubey J, Schares G, and Ortega-Mora L. 2007. Epidemiology and control of neosporosis and *Neospora caninum*. Clin Microbiol Rev 20: 323-367.
- Ekşi F, Demir P, Babür C, and Ütük AE. 2018. A Research on the Seroprevalence of *Toxoplasma gondii* and *Neospora caninum* in Sheep in Adana Province of Turkey. Etlik Vet Mikrobiyol Derg 29: 19-23.
- Ezatpour B, Alirezaei M, Hassanvand A, Zibaei M, Azadpour M, and Ebrahimzadeh F. 2015. The first report of *Neospora caninum* prevalence in aborted and healthy sheep from west of Iran. Comp Clin Path 24: 19-22.
- Figliuolo LPC, Rodrigues A, Viana R, Aguiar DMd, Kasai N, and Gennari SM. 2004. Prevalence of anti-*Toxoplasma gondii* and anti-*Neospora caninum* antibodies in goat from São Paulo State, Brazil. Small Rumin Res 55: 29-32.
- Fish L, Mazuz M, Molad T, Savitsky I, and Shkap V. 2007. Isolation of *Neospora caninum* from dairy zero grazing cattle in Israel. Veterinary parasitology 149: 167-171.
- Gharekhani J, Esmaeilnejad B, Rezaei H, Yakhchali M, Heidari H, and Azhari M. 2016. Prevalence of anti-*Neospora caninum* antibodies in Iranian goats. Ann Parasitol 62.
- Gökçe G, Mor N, Kırmizigul A, Bozukluhan K, and Erkılıc E. 2015. The first report of seropositivity for *Neospora caninum* in sheep from Turkey. Isr J Vet Med 70: 40-44.
- Har U, and Başbuğan Y. 2019. Seroprevalence of *Neospora* caninum in Aborted Sheep in Gevaş District of Van Province. Van Sag Bil Derg 12: 6-12.
- Hässig M, Sager H, Reitt K, Ziegler D, Strabel D, and Gottstein B. 2003. *Neospora caninum* in sheep: a herd case report. Vet parasitol 117: 213-220.
- Howe L, West D, Collett M, Tattersfield G, Pattison R, Pomroy W, Kenyon P, Morris S, and Williamson N. 2008. The role of *Neospora caninum* in three cases of unexplained ewe abortions in the southern North Island of New Zealand. Small Rumin Res 75: 115-122.
- Hughes J, Williams R, Morley E, Cook D, Terry R, Murphy R, Smith J, and Hide G. 2006. The prevalence of *Neospora caninum* and co-infection with *Toxoplasma gondii* by PCR analysis in naturally occurring mammal populations. Parasitol 132: 29-36.
- Hurtado A, Aduriz G, Moreno B, Barandika J, and García-Pérez AL. 2001. Single tube nested PCR for the detection of *Toxoplasma gondii* in fetal tissues from naturally aborted ewes. Vet parasitol 102: 17-27.

- Ibrahim HM, Mohamed AH, El-Sharaawy AA, and El-Shqanqery HE. 2017. Molecular and serological prevalence of *Toxoplasma gondii* in pregnant women and sheep in Egypt. Asian Pac J Trop Med 10: 996-1001.
- Innes EA, Bartley PM, Buxton D, and Katzer F. 2009. Ovine toxoplasmosis. Parasitol 136: 1887-1894.
- Innes EA, Lunden A, Esteban I, Marks J, Maley S, Wright S, Rae A, Harkins D, Vermeulen A, and McKendrick I. 2001. A previous infection with *Toxoplasma gondii* does not protect against a challenge with *Neospora caninum* in pregnant sheep. Parasite Immunol 23: 121-132.
- Koyama T, Kobayashi Y, Omata Y, Yamada M, Furuoka H, Maeda R, Matsui T, Saito A, and Mikami T. 2001. Isolation of *Neospora caninum* from the brain of a pregnant sheep. J Parasitol 87: 1486-1488.
- Mor N, and Arslan MÖ. 2007. Seroprevalence of *Toxoplasma* gondii in Sheep in Kars Province. Kafkas Üniv Vet Fak Derg 13: 165-170.
- Moreno B, Collantes-Fernández E, Villa A, Navarro A, Regidor-Cerrillo J, and Ortega-Mora L. 2012. Occurrence of *Neospora caninum* and *Toxoplasma gondii* infections in ovine and caprine abortions. Vet parasitol 187: 312-318.
- Muz MN, Altuğ N, and Karakavuk M. 2013. Seroprevalence of T. gondii in Dairy Ruminant Production Systems, Shepherd Dogs among the Herds and Detection of *T. gondii*-like Oocyst in Cat Feces in Hatay Region. AVKAE Derg 3: 38-45.
- Nasir A, Ashraf M, Khan M, Javeed A, Yaqub T, Avais M, and Reichel M. 2012. Prevalence of *Neospora caninum* antibodies in sheep and goats in Pakistan. J Parasitol 98: 213-215.
- Oncel T, Vural G, Babur C, and Kilic S. 2005. Detection of toxoplasmosis gondii seropositivity in sheep in Yalova by Sabin Feldman dye test and latex agglutination test. Turkiye Parazitol Derg 29: 2.
- Orunç Kılınç Ö, Ayan A, Yumuşak N, Kömüroğlu AU, Aslan B, Çelik ÖY, and Göz Y. 2023. Investigation of *Toxoplasma gondii* and *Neospora caninum* in different tissues of aborted foetuses of sheep in Van Province, Türkiye: Analysis by nested PCR, histopathological and immunohistochemical methods. Acta Veterinaria Brno 92: 123-131.
- Özkaraca M, İrehan B, Parmaksiz A, Ekinci Aİ, and Çomakli S. 2016. Determination of *Neospora caninum* and *Toxoplasma gondii* in Sheep and Goat Fetuses Using Dublex PCR, Immunohistochemistry, and Immunflourescence Methods. Atatürk Üniversitesi Vet Bil Derg 11: 200-206.
- Öztürk C, Babür C, and Aslan G. 2002. Investigation of antitoxoplasma gondii antibodies in the workers of the slaughter house and the sheep existing in Mersin region. Genel Tıp Dergisi 12: 21-25.

- Panadero R, Painceira A, López C, Vázquez L, Paz A, Díaz P, Dacal V, Cienfuegos S, Fernández G, and Lago N. 2010. Seroprevalence of *Toxoplasma gondii* and *Neospora caninum* in wild and domestic ruminants sharing pastures in Galicia (Northwest Spain). Res Vet Sci 88: 111-115.
- Partoandazanpoor A, Sadeghi-Dehkordi Z, Ekradi L, Khordadmehr M, Rassouli M, and Sazmand A. 2020. Molecular diagnosis and pathological study of *Toxoplasma gondii* in aborted caprine and ovine fetuses in borderline of Iran–Iraq. Acta Parasitol 65: 187-192.
- Raeghi S, Akaberi A, and Sedeghi S. 2011. Seroprevalence of *Toxoplasma gondii* in sheep, cattle and horses in Urmia North-West of Iran. Iran J Parasitol 6: 90-94.
- Sah RP, Dey AR, Rahman AA, Alam MZ, and Talukder MH. 2019. Molecular detection of *Toxoplasma gondii* from aborted fetuses of sheep, goats and cattle in Bangladesh. Veterinary Parasitology: Regional Studies and Reports 18: 100347.
- Shahbazi G, Rad NH, Madani R, Matin S, Mortazavi P, and Jangjou AH. 2019. *Toxoplasma gondii* in aborted fetuses of sheep in Ardebil Area, North-West of Iran. Iran J Parasitol 14: 430-435.
- Sharma RN, Bush J, Tiwari K, Chikweto A, and Bhaiyat MI. 2015. Seroprevalence of *Neospora caninum* in sheep and goats from Grenada, West Indies. Open J. Vet. Med 5: 219.
- Sharma S, Sandhu K, Bal M, Kumar H, Verma S, and Dubey J. 2008. Serological survey of antibodies to *Toxoplasma gondii* in sheep, cattle, and buffaloes in Punjab, India. J Parasitol 94: 1174-1175.
- Tenter AM, Heckeroth AR, and Weiss LM. 2000. *Toxoplasma gondii*: from animals to humans. Int J Parasitol 30: 1217-1258.
- Ueno TEH, Gonçalves VSP, Heinemann MB, Dilli TLB, Akimoto BM, de Souza SLP, Gennari SM, and Soares RM. 2009. Prevalence of *Toxoplasma gondii* and *Neospora caninum* infections in sheep from Federal District, central region of Brazil. Trop Anim Health Prod 41: 547-552.
- Uzêda RS, Pinheiro AM, Fernández SY, Ayres MCC, Gondim LFP, and Almeida MAOd. 2007. Seroprevalence of *Neospora caninum* in dairy goats from Bahia, Brazil. Small Rumin Res 70: 257-259.
- Van der Puije W, Bosompem K, Canacoo E, Wastling J, and Akanmori B. 2000. The prevalence of anti-*Toxoplasma* gondii antibodies in Ghanaian sheep and goats. Acta Tropica 76: 21-26.
- Wang C, Qiu J, Gao J, Liu L, Wang C, Liu Q, Yan C, and Zhu X. 2011. Seroprevalence of *Toxoplasma gondii* infection in sheep and goats in northeastern China. Small Rumin Res 97: 130-133.